

Motivation and Objectives

The development of colloidal lignin particles (CLPs) is a promising strategy to overcome the challenges of lignin application, such as its poor water solubility [1]. The antisolvent precipitation is a widely used technique to produce CLPs, presenting several advantages, however, studies using this technique present some drawbacks.

Advantages

Simple

High particle yield

Controllable size and morphology

Low cost

Drawbacks

Low particle concentration

Toxic solvents

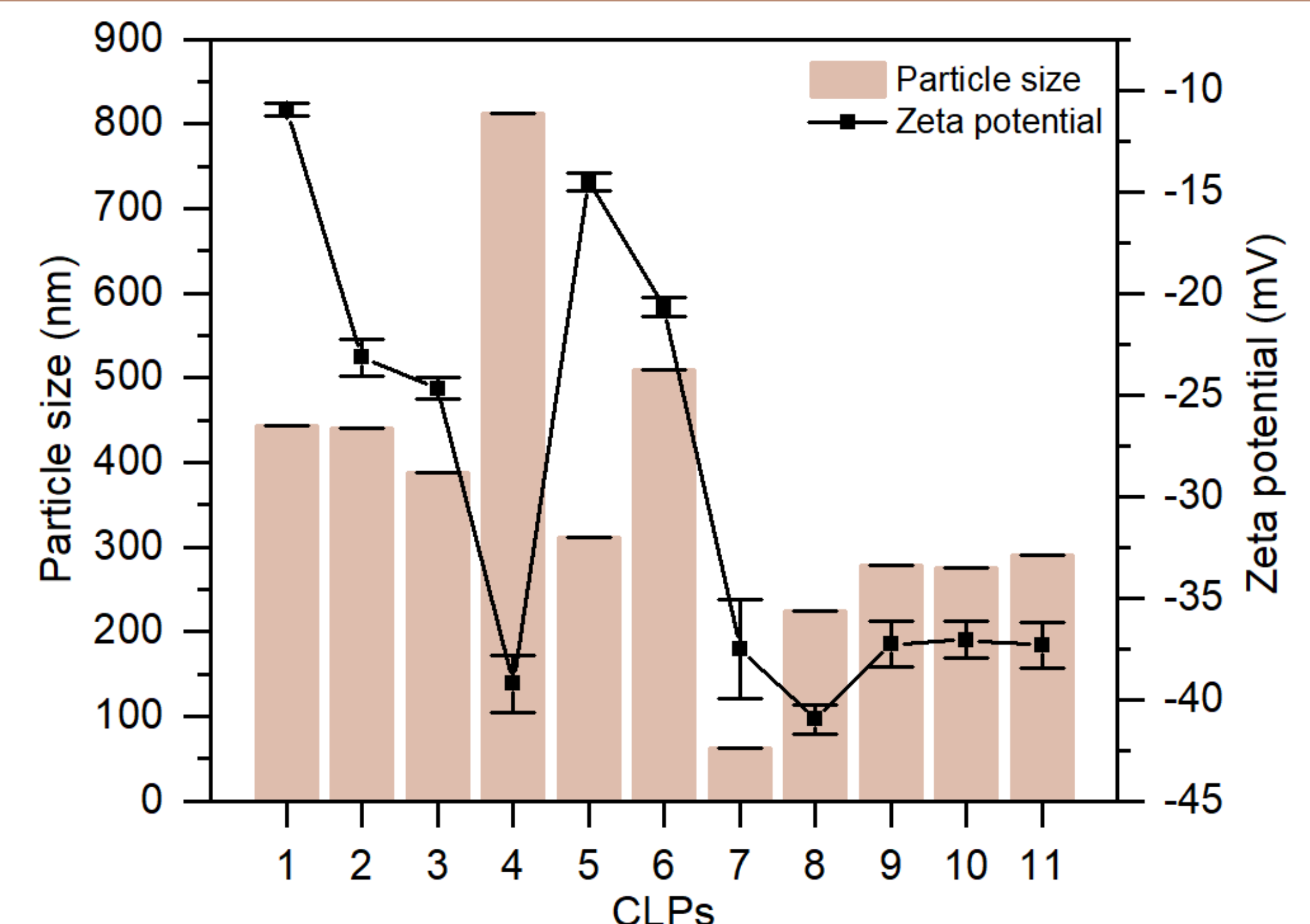


To produce concentrated and stable CLPs through the antisolvent precipitation technique



Increase knowledge on process variables and their effects on particle characteristics

Particle Size and Zeta Potential

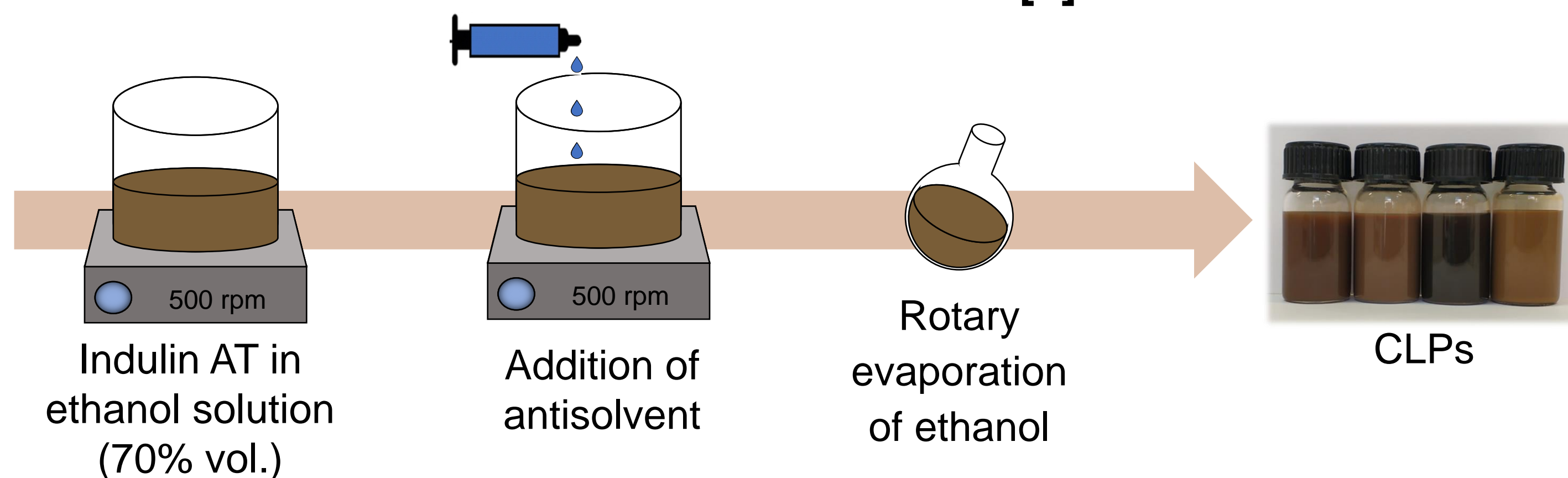


| Effect Analysis (FFD 2 ⁴⁻¹) | Particle size | | Zeta potential | |
|---|---------------|----------------|----------------|---------------|
| | Effect (nm) | p (<0.10) | Effect (mV) | p (<0.10) |
| Mean | 398.99 | 0.0006 | -26.46 | 0.0000 |
| Curvature | -234.64 | 0.2952 | -21.54 | 0.0057 |
| Initial lignin concentration (10 to 50 g/L) | ↑196.03 | 0.1205* | ↓-9.10 | 0.0135 |
| Antisolvent pH (4 to 8) | -54.53 | 0.6252 | ↓-18.27 | 0.0007 |
| Final ethanol concentration (15 to 45%) | ↓-244.03 | 0.0674 | -3.91 | 0.1689 |
| Antisolvent addition rate (6 to 14 mL/min) | -115.48 | 0.3209 | 1.26 | 0.6259 |

*Considering the proximity of 0.1205 with the p-value significance level of 0.10 (10%), the initial lignin concentration was considered a significant factor for the particle size response.

Methodology

Production of CLPs [1]



Study of process variables effects Fractional Factorial Design (FFD) 2⁴⁻¹

| Assay | Decodified values | | | |
|-------|------------------------------------|----------------|--|------------------------------------|
| | Initial lignin concentration (g/L) | Antisolvent pH | Final ethanol concentration (% volume) | Antisolvent addition rate (mL/min) |
| 1 | 10 | 4 | 15 | 6 |
| 2 | 50 | 4 | 15 | 14 |
| 3 | 10 | 8 | 15 | 14 |
| 4 | 50 | 8 | 15 | 6 |
| 5 | 10 | 4 | 45 | 14 |
| 6 | 50 | 4 | 45 | 6 |
| 7 | 10 | 8 | 45 | 6 |
| 8 | 50 | 8 | 45 | 14 |
| 9 | 30 | 6 | 30 | 10 |
| 10 | 30 | 6 | 30 | 10 |
| 11 | 30 | 6 | 30 | 10 |

Responses

- Particle size (Laser Diffraction Analysis, D50 number-based distribution)
- Zeta potential (Nano-ZS Zetasizer)
- Color (Colorimeter CR-400 Konica Minolta, L*a*b* CIELAB space)
- Emulsifying potential (Mixture of CLPs and Miglyol 812, vortex for 1 minute). Emulsified Layer (EL %) calculated as Equation 1:

$$EL\% = \frac{Emulsion_{height}}{Total_{height}} \times 100 \quad (1)$$

Conclusions and Future Work

- The **antisolvent pH** and **initial lignin concentration** were **significant variables** affecting the stability, size, and color of CLPs.
- It was possible to produce **stable CLPs** at a concentration up to **50 g/L**
- CLPs** presented **high potential** as **Pickering emulsions stabilizers**

Ongoing/Future work: production of Pickering emulsions using CLPs as stabilizers for bio-based applications.

Color of CLPs

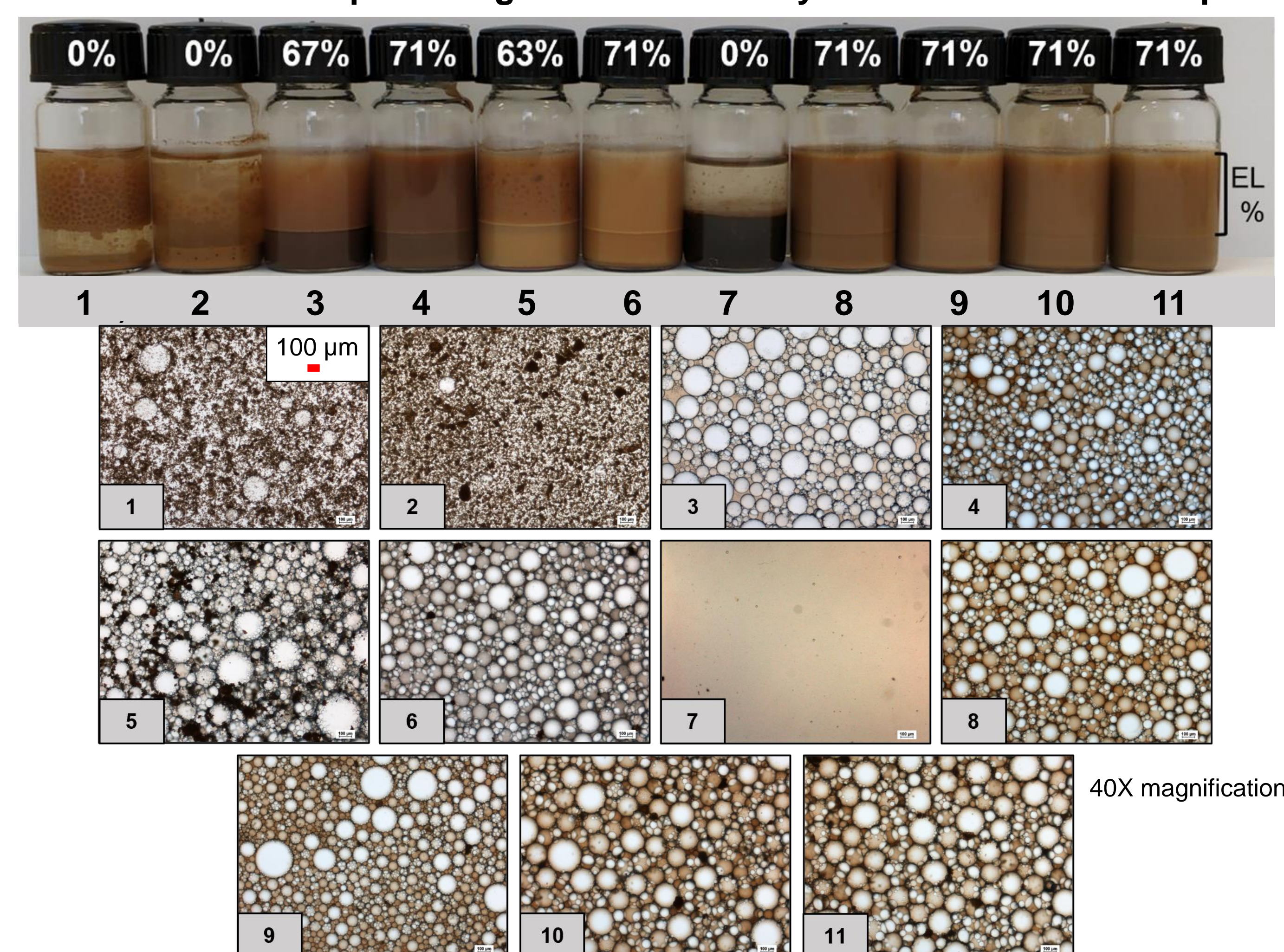


- ↑ Initial lignin concentration (10 to 50 g/L) = ↑ Luminosity and yellowish tendency
- ↑ Antisolvent pH (4 to 8) = ↓ Luminosity ↑ brownish tendency

Emulsifying Potential

Mixtures of CLPs and Miglyol 812 (50/50 vol.)

EL% indicates the percentage of emulsified layer formed for each sample



- Only CLPs 1, 2 and 7 were not able to form an emulsified layer, due to particle instability (CLPs 1 and 2) or low particle size and concentration (CLPs 7)

References and Acknowledgments

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References. [1] Colucci, G., Santamaria-Echart, A., Silva, S. C., Teixeira, L. G., Ribeiro, A., Rodrigues, A. E., Barreiro, M. F. Colloids Surf. A: Physicochem. Eng. Asp., **666** (2023) 131287.